

## Hydroxyproline assay by HPLC-FLD applied for wound healing determination in rat model

Silvia Kociova<sup>1,2</sup>, Zuzana Lackova<sup>1,2</sup>, Natalia Cernei<sup>1,2</sup>, Dagmar Sterbova<sup>1</sup>,  
Tomas Komprda<sup>3</sup>, Ondrej Zitka<sup>1,2,4</sup>

<sup>1</sup>Department of Chemistry and Biochemistry  
Mendel University in Brno

<sup>2</sup>Central European Institute of Technology  
Brno University of Technology  
Purkynova 123, 612 00 Brno

<sup>3</sup>Department of Food Technology

<sup>4</sup>CEITEC – Central European Institute of Technology  
Mendel University in Brno  
Zemedelska 1, 613 00 Brno  
CZECH REPUBLIC

xkociova@mendelu.cz

**Abstract:** In this work, we focused on four types of oils and their response on wound healing of the skin of rats after dietary exposure. A set of samples consisted of 48 adult male rats of *Wistar albino*, which were divided into four groups according to the respective oil. The content of hydroxyproline in the skin tissue, used as a biomarker for wound healing, was determined by high-performance liquid chromatography with a fluorescence detector. There was found out, that the highest content of hydroxyproline was detected in the group of rats, which have been fed with the addition of palm oil ( $118.4 \pm 1.9 \mu\text{g/g}$ ). These results indicate that palm oil had the most positive effect on wound healing.

**Key Words:** high-performance liquid chromatography with fluorescence detector (HPLC-FLD), hydroxyproline, microwave hydrolysis, rat skin

### INTRODUCTION

Hydroxyproline is a major part of the collagen of animals (Zhang and Duan 2017) and human (Sugioka et al. 2017) and it is responsible for collagen stability (Shoulders and Raines 2009). Furthermore, the presence of hydroxyproline and its concentration in biological fluids can be used as a marker of collagen catabolism and tissue degradation or bone resorption. Various changes in hydroxyproline metabolism play major role in the pathogenesis of different diseases (Srivastava et al. 2016). The higher level of hydroxyproline is observed in cases of depression and stress, or in cases of poor wound-healing (Srivastava et al. 2016, Dong et al. 2017). In the case of diagnosed fibrosis in hepatitis C, adiposity and cardiometabolic health, hydroxyproline may be used as potential oxidative biomarker. Nowadays, a high-performance liquid chromatography with fluorescence detection, the high-performance liquid chromatography with tandem mass spectrometric detection (LC-MS/MS), a capillary electrophoresis with in-capillary optical fiber light-emitting diode-induced fluorescence detection (Ji et al. 2018) and a hydrophilic interaction chromatography-quadrupole/electrostatic field orbitrap high resolution mass spectrometry (Liu et al. 2017) are used for determination of hydroxyproline in tissues or body fluids.

This work was focused on determination of hydroxyproline in rat's skin using a high-performance liquid chromatography with fluorescence detector. Figure 1 demonstrates the metabolism of hydroxyproline in animals.

The diagram illustrates the metabolic pathway of 4-Hydroxyproline degradation. The pathway begins with 4-Hydroxyproline, which is converted to  $\Delta^1$ -Pyrroline-3-hydroxy-5-carboxylate by the enzyme 4-Hydroxylase, using FAD as a cofactor and producing FADH<sub>2</sub>. This intermediate is then converted to 4-erythro-hydroxy-glutamate by 3-OH+P5C dehydrogenase, using NAD<sup>+</sup> as a cofactor and producing NADH+H<sup>+</sup>. 4-erythro-hydroxy-glutamate is converted to 4-hydroxy-2-ketoglutarate by GOT, using OAA as a cofactor and producing Glu. 4-hydroxy-2-ketoglutarate is then converted to Glyoxylate by 4-OH-2-KG aldolase. Glyoxylate is converted to Oxalate by LDH, using Ala as a cofactor and producing Pyruvate. Oxalate is excreted in urine. Glyoxylate is also converted to Glycine by ATG, using Pyr as a cofactor and producing Pyruvate. Glycine is converted to Glucose by DAO. Glucose is converted to Acetyl-CoA by PDH, producing CO<sub>2</sub>. Acetyl-CoA is then converted to ATP by the TCA cycle, producing CO<sub>2</sub> and H<sub>2</sub>O. 4-Hydroxyproline is also converted to  $\Delta^1$ -Pyrroline-4-hydroxy-2-carboxylate by LAA-oxidase, which is then converted to Pyrrole-2-carboxylate spontaneously. Pyrrole-2-carboxylate is excreted in urine.

580 | Page

size of 3.5  $\mu\text{m}$  was used. The column was equilibrated at 40 °C. The mobile phases A consisted of 40 mM sodium phosphate dibasic at pH 7.8 and mobile phase B acetonitrile/methanol/water (45:45:10 v/v/v). The flow rate of the mobile phase was 2 ml/min. The compounds were eluted with a linear upward gradient 0.0 min (0% B)  $\rightarrow$  1.9 min (0% B)  $\rightarrow$  18.1 min (57% B)  $\rightarrow$  18.6 min (100% B)  $\rightarrow$  22.3 min (100% B)  $\rightarrow$  23.2 min (0% B).

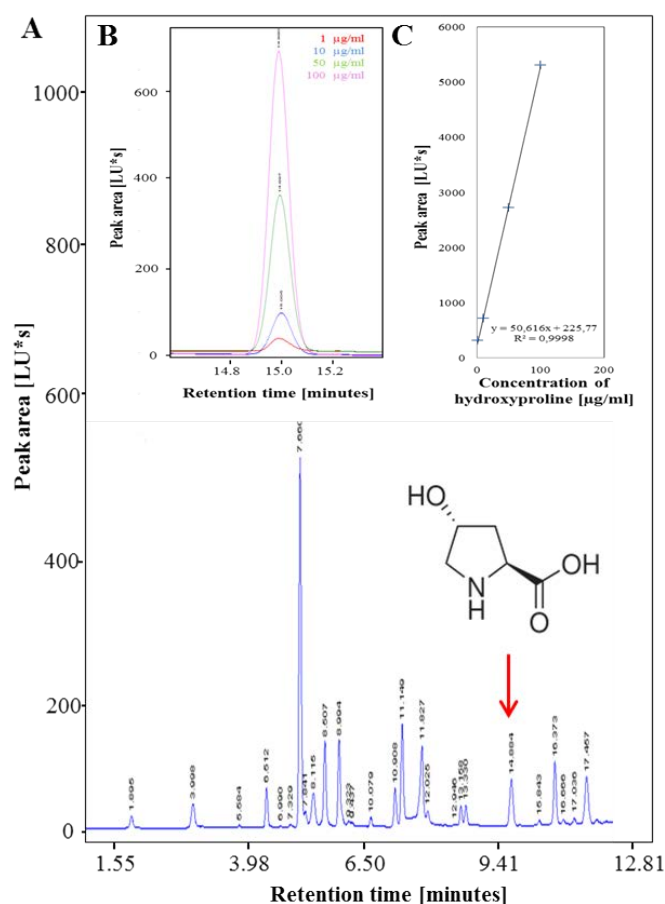
### Statistical analyses

The detection limits (3 signal/noise, S/N) were calculated according to Long and Winefordner (Long and Winefordner 1983), whereas N was expressed as standard deviation of noise determined in the signal domain unless stated otherwise.

## RESULTS AND DISCUSSION

The determination of the presence and of the concentration of hydroxyproline in rat skin samples was done by high-performance liquid chromatography with fluorescence detection (HPLC-FLD). The content of hydroxyproline in a rat skin was expressed as a mean  $\pm$  standard deviation from three replicates. Calibration curve was prepared in the range 1–100  $\mu\text{g/ml}$  for HPLC-FLD. For analysis of hydroxyproline, the limit of detection (LOD) was 0.33 ng/ml, whereas the limit of quantification (LOQ) was 1.00 ng/ml. The calibration curve showed a good linearity with correlation coefficient  $R^2 = 0.9998$  and R.S.D. = 1.5% (Figure 2 B, C). The figure 2A shows the HPLC-FLD chromatographic analysis record of real samples of rat skin.

Figure 2 (A) Chromatographic record of a real sample of rat skin, (B) chromatograms from calibration dependence of hydroxyproline determined by HPLC-FLD, (C) calibration curve measured within the range from 1 to 100  $\mu\text{g/ml}$  under the experimental conditions.

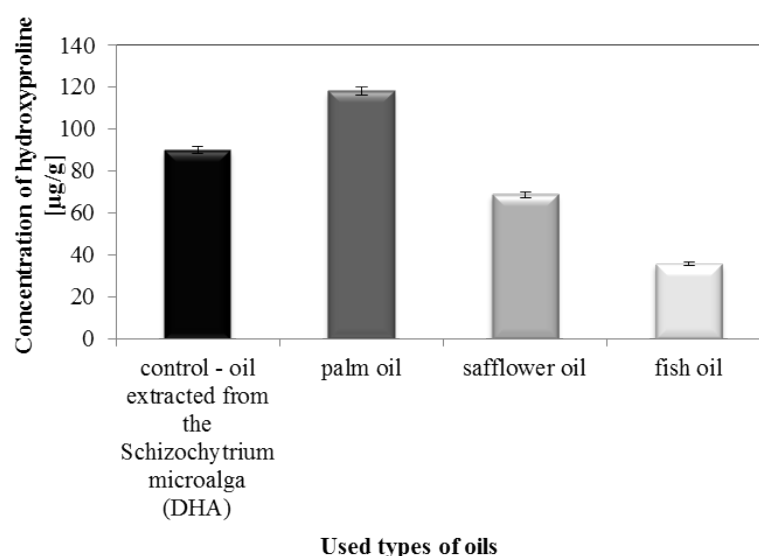


Sample analysis was performed by calibrated high-performance liquid chromatography with fluorescence detection (HPLC-FLD). As a control, an oil extracted from the *Schizochytrium microalga* (DHA) was taken up with a value of  $90.2 \pm 1.7 \mu\text{g/g}$ . In Figure 3 is obvious, that the highest

content of hydroxyproline was recorded in samples of rats, which have been fed with palm oil ( $118.4 \pm 1.9 \mu\text{g/g}$ ) compared to other oil supplements. The results indicate that the most positive effect on wound healing had a palm oil. Results of content of hydroxyproline for safflower oil ( $68.9 \pm 1.3 \mu\text{g/g}$ ) and fish oil ( $35.7 \pm 0.9 \mu\text{g/g}$ ) are lower than control results.

Only one third of hydroxyproline content in wound was determined against the control group after 18 days after excision of rats fed with fish oil ( $36 \mu\text{g/g}$  vs.  $118 \mu\text{g/g}$ ; Figure 3). This result agrees with the data of Gercek and co-workers (Gercek et al. 2007) who reported lower hydroxyproline content ( $99.3\text{--}99.8 \mu\text{g/g}$ ) compared to control (saline;  $152.9 \mu\text{g/g}$ ) in wounds of rats treated five days after wounding with parenteral fish oil emulsion. Similarly, Rosa and co-workers (Rosa et al. 2014) found out lower hydroxyproline content (ca  $0.25 \mu\text{g/g}$ ) fourteen days post-excision in wounds of mice ingesting (by gavage) fish oil in comparison with both olive oil (ca  $0.50 \mu\text{g/g}$ ; high content of 18:1 n-9 oleic acid) and a control (water; ca  $1 \mu\text{g/g}$ ). Otranto and co-workers (Otranto et al. 2010) fed rats thirty days before cutaneous wounding with a diet supplemented with (among others) a fish oil and sunflower oil and found out two times higher hydroxyproline content in wounds fourteen days later in the fish oil group (about  $2.2 \mu\text{g/g}$ ) compared to the sunflower oil group (ca  $1.1 \mu\text{g/g}$ ).

*Figure 3 Hydroxyproline content 18 days post-excision in wounded skin of rats fed with a diet supplemented with 5% of Schizochytrium microalga extract (control), 5% of palm oil, 5% of safflower oil and 5% of fish oil.*



*Legend: The values are expressed as the mean of twelve independent replicates ( $n=12$ ). Vertical bars indicate standard error  $p<0.05$ .*

All the above-mentioned experiments of (Gercek et al. 2007), (Rosa et al. 2014) and (Otranto et al. 2010), used a “basic” spectrophotometric determination according to (Woessner 1961), while HPLC method was applied in this work. Moreover, considering the fact that collagen fibrils begin to appear by the fifth day and collagen continues to accumulate within the second week (Hashimoto et al. 2002), the absolute amounts of hydroxyproline measured at different time intervals after the excision are practically incomparable. Only a relative comparison of dietary interventions is meaningful.

## CONCLUSION

The aim of this experiment was to determine the concentration of hydroxyproline in the rats' skin wound after supplementation with 5% of palm oil, 5% of safflower oil, 5% of fish oil and 5% of oil extracted from the *Schizochytrium microalga* (marked as DHA; control) in the feed. The results showed the highest concentration of hydroxyproline in samples of rats that have been fed with palm oil ( $118.4 \pm 1.9 \mu\text{g/g}$ ) compared to the others. The results suggest that the most positive effect on wound healing had palm oil. Content of hydroxyproline for safflower oil ( $68.9 \pm 1.3 \mu\text{g/g}$ ) and fish oil

( $35.7 \pm 0.9 \mu\text{g/g}$ ) is lower than control ( $90.2 \pm 1.7 \mu\text{g/g}$ ). This data will serve as a pilot study for a larger experiment.

## ACKNOWLEDGEMENTS

This research was carried out under the project of the Internal Grant Agency of Mendel University in Brno (AF-IGA2019-TP006) and by the project CEITEC 2020 (LQ1601) with financial support from the Ministry of Education, Youth and Sports of the Czech Republic under the National Sustainability Programme II and by EFRR “Multidisciplinary research to increase application potential of nanomaterials in agricultural” (No. CZ.02.1.01/0.0/0.0/16\_025/0007314).

## REFERENCES

- Dong, W.W. et al. 2017. Protective Effects of Hydrogen-Rich Saline Against Lipopolysaccharide-Induced Alveolar Epithelial-to-Mesenchymal Transition and Pulmonary Fibrosis. *Medical Science Monitor* [Online], 23: 2357–2364. Available at: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5445901/>. [2018-08-17].
- Gercek, A. et al. 2007. Effects of parenteral fish-oil emulsion (Omegaven) on cutaneous wound healing in rats treated with dexamethasone. *Journal of Parenteral and Enteral Nutrition* [Online], 31(3): 161–166. Available at: <https://onlinelibrary.wiley.com/doi/abs/10.1177/0148607107031003161>. [2018-08-17].
- Hashimoto, I. et al. 2002. Angiostatic effects of corticosteroid on wound healing of the rabbit ear. *Journal of Medical Investigation* [Online], 49(1-2): 61–66. Available at: <https://www.ncbi.nlm.nih.gov/pubmed/11901762>. [2018-08-17].
- Ji, H.Y. et al. 2018. Sensitive determination of l-hydroxyproline in dairy products by capillary electrophoresis with in-capillary optical fiber light-emitting diode-induced fluorescence detection. *Analytical Methods* [Online], 10(19): 2211–2216. Available at: <https://pubs.rsc.org/en/content/articlelanding/2017/ay/c7ay02356a/unauth#!divAbstract>. [2018-08-17].
- Liu, W. et al. 2017. Determination of hydroxyproline in liver tissue by hydrophilic interaction chromatography-quadrupole / electrostatic field orbitrap high resolution mass spectrometry. *Chinese Journal of Chromatography* [Online], 35(12): 1251–1256. Available at: <https://www.ncbi.nlm.nih.gov/pubmed/29372775>. [2018-08-17].
- Long, G.L., Winefordner J.D. 1983. Limit of Detection A Closer Look at the IUPAC Definition. *Analytical Chemistry* [Online], 55(07): 712A–724A. Available at: <https://pubs.acs.org/doi/pdf/10.1021/ac00258a724>. [2018-08-17].
- Otranto, M. et al. 2010. Effects of supplementation with different edible oils on cutaneous wound healing. *Wound Repair and Regeneration* [Online], 18(6): 629–636. Available at: <https://onlinelibrary.wiley.com/doi/abs/10.1111/j.1524-475X.2010.00617.x>. [2018-08-17].
- Rosa, A.D.S. et al. 2014. Supplementation with olive oil, but not fish oil, improves cutaneous wound healing in stressed mice. *Wound Repair and Regeneration* [Online], 22(4): 537–547. Available at: <https://onlinelibrary.wiley.com/doi/abs/10.1111/wrr.12191>. [2018-08-17].
- Shoulders, M.D., Raines R.T. 2009. Collagen Structure and Stability. *Annual Review of Biochemistry*. Palo Alto, Annual Reviews [Online], 78: 929–958. Available at: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2846778/>. [2018-08-17].
- Srivastava, A.K. et al. 2016. Hydroxyproline: A Potential Biochemical Marker and Its Role in the Pathogenesis of Different Diseases. *Current Protein & Peptide Science* [Online], 17(6): 596–602. Available at: <http://www.eurekaselect.com/137423/article>. [2018-08-17].
- Sugioka, K. et al. 2017. Extracellular Collagen Promotes Interleukin-1 beta-Induced Urokinase-Type Plasminogen Activator Production by Human Corneal Fibroblasts. *Investigative Ophthalmology & Visual Science* [Online], 58(3): 1487–1498. Available at: <https://iovs.arvojournals.org/article.aspx?articleid=2610161>. [2018-08-17].
- Woessner Jr, J.F. 1961. The determination of hydroxyproline in tissue and protein samples containing small proportions of this imino acid. *Archives of Biochemistry and Biophysics* [Online], 93(2): 440–447. Available at: <https://www.sciencedirect.com/science/article/pii/0003986161902910>. [2018-08-17].

- Wu, G.Y. et al. 2011. Proline and hydroxyproline metabolism: implications for animal and human nutrition. *Amino Acids* [Online], 40(4): 1053–1063. Available at: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3773366/>. [2018-08-17].
- Zhang, J.J., Duan R. 2017. Characterisation of acid-soluble and pepsin-solubilised collagen from frog (*Rana nigromaculata*) skin. *International Journal of Biological Macromolecules* [Online], 101: 638–642. Available at: <https://www.sciencedirect.com/science/article/pii/S0141813017304002?via%3Dihub>. [2018-08-17].